We describe the first isolation of an NDM-1-producing *Pseudomonas aeruginosa* in Italy. In May 2013, a patient with acute lymphoblastic leukaemia and history of prior hospitalisation in Belgrad, Serbia, underwent stem cell transplantation at a tertiary care hospital in Rome, Italy. After transplantation, sepsis by NDM-1-producing *P. aeruginosa* occurred, leading to septic shock and fatal outcome.

In May 2013, a *Pseudomonas aeruginosa* strain producing New Delhi metallo-beta-lactamase-1 (NDM-1) was isolated in Italy from a patient previously hospitalised in Serbia and found to be resistant to all antibiotics tested except colistin. Multidrug-resistant and extensively drug-resistant *P. aeruginosa* represent an increasing therapeutic challenge worldwide. In particular, the ST235 clone is the founder of the successful epidemic clonal complex 235 (CC235), associated with various carbapenemase genes, but very rarely with NDM-1 [1,2].

**Case description**

In late May 2013, a man in his early 40s with a diagnosis of acute lymphoblastic leukaemia (ALL) in first remission was admitted to the haematology unit of a hospital in Rome, Italy, in order to undergo stem cell transplantation. He had been hospitalised in December 2012 at a general hospital in Belgrad, Serbia. In that hospital, the patient had undergone four courses of chemotherapy (HYPER-C-VAD) from December 2012 to February 2013 with a complete remission of disease.

At hospital admission in Rome, the patient was febrile, in good general condition. According to the haematology ward’s routine procedure adopted for transplantation candidates, nasal, pharyngeal, rectal and groin swabs, as well as blood, urine and stool cultures were screened and found negative for multidrug-resistant bacteria. In brief, all samples were screened for methicillin-resistant *Staphylococcus aureus* (MRSA), extended-spectrum beta-lactamase (ESBL)-producing *Enterobacteriaceae*, *Acinetobacter baumannii*, *P. aeruginosa*, carbapenem-resistant *Enterobacteriaceae*, vancomycin-resistant *Enterococcus faecium* (VRE), using MacConkey and chromogenic agar plates (Becton Dickinson, Heidelberg, Germany). Antimicrobial drug susceptibility testing and minimal inhibitory concentrations (MICs) were obtained by the Phoenix System (Becton Dickinson, Sparks, MD) and interpreted according to the recommendations from the European Committee on Antimicrobial Susceptibility Testing (EUCAST) [3].

Lacking an available HLA-compatible donor, the patient received on 28 May a stem cell transplant from a haploidentical donor, with a high-dose conditioning regimen containing thiopeta, busulfan, and also containing fludarabine-unmanipulated bone marrow as the stem cell source. Prophylaxis of graft-versus-host disease was post-transplant cyclophosphamide, mycophenolate and cyclosporine, as in the Baltimore programme [4]. Two days later he developed severe neutropenia (absolute neutrophil count (ANC) =0.1x10^9/L) and treatment was started with piperacillin/tazobactam, amikacin and vancomycin, with an immediate defervescence. The patient remained neutropenic (ANC <0.1x10^9/L), and fever reappeared some days later. Meropenem was added in substitution of piperacillin/tazobactam.

A pharyngeal swab taken 10 days after transplantation was positive for *Klebsiella pneumoniae*. Species identification and antimicrobial susceptibilities were determined by the Phoenix System (Becton Dickinson, Sparks, MD) and the tigecycline MIC was obtained by Etest (Becton-Dickinson, United States). The *K. pneumoniae* isolate was an ESBL producer with resistance to cefotaxime (MIC >4 mg/L) ceftazidime (MIC >8 mg/ml), ertapenem (MIC >1 mg/L) and tigecycline.
At Day 15 after transplantation, the patient was still febrile and neutropenic. Another rectal swab was again positive for the ertapenem-resistant *K. pneumoniae*. A perianal abscess appeared on Day 16 and draining yielded *P. aeruginosa* and VRE. This isolate was resistant to all antibiotics tested (Table) except colistin.

The blood cultures of the patient, taken on Day 15, yielded a *P. aeruginosa* with an identical phenotype. Vancomycin treatment was halted, and tigecycline (200 mg/day administered every 12 hours, after a loading dose of 200 mg), rifampicin (600 mg given every 24 hours) and daptomycin (700 mg every 24 hours), were added to the previous treatment.

On Day 17 after transplantation, the patient rapidly deteriorated and was transferred to the intensive care unit, where he died few hours later from septic shock still in profound aplasia.

### Table

<table>
<thead>
<tr>
<th>Antibiotics</th>
<th>MIC (µg/mL), Interpretation result†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amikacin</td>
<td>&gt;16, R</td>
</tr>
<tr>
<td>Aztreonam</td>
<td>=16, R</td>
</tr>
<tr>
<td>Ceftazidime</td>
<td>&gt;8, R</td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>&gt;1, R</td>
</tr>
<tr>
<td>Colistin</td>
<td>≤1, S</td>
</tr>
<tr>
<td>Gentamicin</td>
<td>&gt;4, R</td>
</tr>
<tr>
<td>Imipenem</td>
<td>&gt;8, R</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>&gt;2, R</td>
</tr>
<tr>
<td>Meropenem</td>
<td>&gt;8, R</td>
</tr>
<tr>
<td>Piperacillin</td>
<td>&gt;16, R</td>
</tr>
<tr>
<td>Piperacillin/tazobactam</td>
<td>&gt;16/4, R</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>&gt;4, R</td>
</tr>
</tbody>
</table>

MIC: minimum inhibitory concentration; R: resistant; S: susceptible

† Based on EUCAST interpretive criteria [1].

(MIC 3 mg/L), but remained susceptible to imipenem (MIC ≤1 mg/L), meropenem (MIC ≤1 mg/L) and colistin (MIC ≤1 mg/L), according to interpretive criteria from the EUCAST [3]. The strain was assigned to ST101 by multilocus sequence typing [5] and resulted negative for carbapenemase genes in PCR using previously described primers and protocols [6, 7]. Colistin (given every 12 hours for a total daily dose of 9,000,000 IU after a loading dose of 9,000,000 IU) was added to the ongoing antimicrobial treatment.

### Molecular characterisation of the *P. aeruginosa* isolate

The *P. aeruginosa* strain isolated from the blood cultures was identified as sequence type (ST) 235 by MLST [8]. The ST235 strain resulted positive for the *bla*<sub>NDM-1</sub> gene. Plasmid and total DNA was tested by Southern blot hybridisation with the *bla*<sub>NDM-1</sub> gene as probe. Results suggested a chromosomal location of the *bla*<sub>NDM-1</sub> gene (data not shown). Total DNA was restricted with HindIII and a genome library was constructed in the HindIII-pZErO-2 kanamycin-resistant vector (Invitrogen, Milan Italy). Transformants were obtained selecting on plates containing 30 mg/L ampicillin and 30 mg/L kanamycin. One transformant carried a HindIII insert of approximately 3.3 KB, which resulted positive in a PCR for the *bla*<sub>NDM-1</sub> gene. The insert was fully sequenced using universal primers and the primer walking approach. Comparative DNA sequence analysis showed that the HindIII insert in our construct had 100% of DNA identity to the HindIII-fragment of the ST235 *P. aeruginosa* strain HIABP11 (EMBL ACC. No. KC170992) carrying *bla*<sub>NDM-1</sub>, that was isolated in France in March 2012 [9]. Based on the DNA sequence of the HIABP11 strain (EMBL ACC No. KC170992), 12,762 kb containing the entire *bla*<sub>NDM-1</sub>-gene environment and flanking regions were analysed by PCR and sequencing. The presence of ISPa7 was identified upstream of the *ndm-1* gene. The insert was fully sequenced using universal primers and the primer walking approach. Comparative DNA sequence analysis showed that the HindIII insert in our construct had 100% of DNA identity to the HindIII-fragment of the ST235 *P. aeruginosa* strain HIABP11 (EMBL ACC. No. KC170992) carrying *bla*<sub>NDM-1</sub>, that was isolated in France in March 2012 [9].

### Control measures

The ST235 strain could represent a serious risk for potential spreading in the hospital environment.

However, during the hospitalisation and after the identification of the strain, strict contact isolation precautions (CIP), and screening of the personnel were implemented on the haematology ward. CIP included wearing a gown and gloves when entering the patient’s room, promptly removing gloves after care and hand decontamination, as well as using disposable single-use or patient dedicated non-critical equipment such as blood pressure cuff and stethoscope. Moreover, since the hands of healthcare workers are the most common vehicle for the transmission of microorganisms, including *P. aeruginosa* and *K. pneumoniae*, from patient to patient and within the healthcare environment [10], hand hygiene practices were strengthened.

All contacts, room-mates, relatives, and personnel involved in the care of the patient were screened for carriage of this organism. In addition, environmental screening for *Pseudomonas* and Klebsiella was performed at several points of the ward, including the rooms where the patient stayed. Neither NDM-1-producing *P. aeruginosa* nor Klebsiella were obtained
from these cultures. Moreover, after a one month of strict surveillance, neither any infection nor any colonization by NDM-1-producing *P. aeruginosa* or *Klebsiella* were observed on the wards where the patient stayed.

**Discussion**

It has been recently demonstrated that the emergence of multidrug-resistant, metallo-beta-lactamase (MBL)-positive *P. aeruginosa* in Russia was largely due to the spread of one dominant clone, CC235 producing VIM-2 [1]. The association of CC235 with MBL genes has been reported outside Russia in several European countries [1]. In particular, VIM-1 was the more common MBL identified in CC235 *P. aeruginosa* from Italy, VIM-4 in Greece, Sweden, Hungary, and Belgium, VIM-13 in Spain, and IMP-29 in France [1]. To the best of our knowledge, only few NDM-1-producing *P. aeruginosa* strains were reported in Europe before this Italian case: two strains were described in Serbia in 2010 and one strain was identified in France in 2012 [2,9]. Both the Italian and French NDM-1 producing *P. aeruginosa* were identified in patients previously hospitalised in Serbian hospitals, where a reservoir of NDM-1-producing CC235 *P. aeruginosa* can be suspected. Moreover, six of 55 travel-associated cases of *Enterobacteriaceae* producing NDM-1 reported in European countries had a link to the Balkan region, presumably to Serbia, Kosovo*, Montenegro, and Bosnia and Herzegovina [11].

* This designation is without prejudice to positions on status, and is in line with United Nations Security Council Resolution 1244/99 and the International Court of Justice Opinion on the Kosovo declaration of independence.

**Acknowledgements**

This work was supported by The Italian FLAGSHIP “InterOmics” project (PB.P05) funded by MIUR and coordinated by the CNR. Renata Galetti's fellowship was supported by the grant 2012/24864-1, Sao Paulo Research Foundation (FAPESP)

**Conflict of interest**

None declared.

**Authors’ contributions**

All authors of this research paper have directly participated in the planning, execution and analysis of this study.

**References**


