Emergence of enterovirus 71 C4a in Denmark, 2009 to 2013

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Enterovirus (EV) 71 has emerged as a primary cause of severe neurologic enterovirus infection in the aftermath of the global polio eradication effort. Eleven subgenotypes of EV71 exist, the C4 subgenotype being associated with large outbreaks in Asia with high mortality rates. This subgenotype has rarely been reported in Europe. In the period between 1 January 2009 and 31 December 2013 a total of 1,447 EV positive samples from 1,143 individuals were sent to the Statens Serum Institute (SSI), and 938 samples from 913 patients were genotyped at the Danish National World Health Organization Reference laboratory for Poliovirus at SSI. Echovirus 6 (Eo6) (n=141 patients), echovirus 30 (E30) (n=114), coxsackievirus A6 (CAo6) (n=96) and EV71 (n=63) were the most prevalent genotypes. We observed a shift in circulating EV71 subgenotypes during the study period, with subgenotype C4 dominating in 2012. A total of 34 EV71 patients were found to be infected with strains of the C4 subgenotype, and phylogenetic analysis revealed that they belonged to the C4a lineage. In our study, the proportions of cases with cerebral and/or sepsis-like symptoms were similar in those affected by C4a (19/34) and those with C1 and C2 (15/35). The majority (n=30) of the 34 EV71 C4 cases were children ≤5 years of age, and males (n=22) were over-represented. Continued EV surveillance is required to monitor the spread of EV71 C4 in Denmark and the rest of Europe.

Introduction

Human enteroviruses (EV) are small, single-stranded RNA-viruses from the Enterovirus genus of the Picornaviridae family. They can cause a range of clinical manifestations from mild mucocutaneous and/or gastrointestinal symptoms, to visceral and severe neurologic diseases with involvement of central nervous system (CNS). Polioviruses used to be the most important EV due to widespread outbreaks of paralytic disease. A rather successful global effort to eradicate polio has now made EV71 the primary cause of severe neurotropic EV-associated infectious diseases [1]. EV71 variants have been classified into three genogroups (GgA, GgB, and GgC), and the latter two are

further subdivided into subgenotypes B1 to B5, and C1 to C5. Currently genogroups B and C are co-circulating worldwide. Subgenotype C1 is predominating in Europe, but it can also be found in Australia, Malaysia and Singapore. The C4 subgenotype has predominantly been identified in large outbreaks of hand, foot and mouth disease (HFMD) in Asia, and in particular mainland China, where severe cases and a rather high mortality rate have been reported [2-4]. In 2004 the C4 subgenotype was detected for the first time in Europe, and has to date only been reported in a total of nine cases in Austria, Croatia, and Hungary, respectively [2-4]. In February 2012, the first EV71 C4 case was detected in Denmark in a Serbian infant admitted to the paediatric ward at Hospital A with fever and CNS symptoms. In the following months more EV71 C4 cases were detected in the same geographical area as the hospital. The Virology Surveillance and Research Section (VOF) at the Department of Microbiological Diagnostics and Virology, Statens Serum Institut (SSI) serves as the National World Health Organization (WHO) Reference Laboratory for Poliovirus in Denmark. The Danish EV surveillance is implemented to monitor poliomyelitis as part of the poliomyelitis elimination efforts in Denmark. We took advantage of the well-functioning EV-surveillance system to characterise the emergence of EV71 C4 strains in Denmark.

Methods

Enterovirus surveillance system

The national EV surveillance system in Denmark is conducted in a joint effort by the National WHO Poliovirus Reference Laboratory at VOF, SSI and the Infectious Disease Epidemiology Department (IDED) at SSI. The system is voluntary and all types of EV positive sample material may be submitted for characterisation. However, as part of the global poliovirus elimination programme, in case a patient is diagnosed with EV in the cerebrospinal fluid (CSF), it is highly requested that CSF and stool are forwarded to the Danish WHO Reference Laboratory for Poliovirus for virus characterisation including culture and viral protein (VP)1/

VP2 sequence-based typing. Samples for diagnostic testing may also be sent directly to SSI from general practitioners and hospitals. Once weekly, VOF, SSI reports the new EV positive cases to the WHO Regional Office for Europe (EURO) in Copenhagen, as well as to the IDED. To ensure that relevant clinical information is archived in the national EV surveillance database, the IDED sends a letter with a standardised questionnaire regarding information on the clinical symptoms (including information on acute flaccid paralysis) and a reminder to send stool for virus characterisation directly to the doctors/departments in charge of the EV-positive patients. Completed questionnaires are returned and data entered in the database by the IDED, SSI. To ensure completeness of clinical data information for this paper, we have contacted relevant hospital departments and asked for relevant information on all EV71 cases where clinical information was missing.

Enterovirus characterisation

Isolates from all severe (i.e. with meningitis, encephalitis and sepsis-like illness) EV positive cases are routinely typed centrally at the VOF at SSI by sequencing part of the VP2 gene from the polymerase chain reaction (PCR) product obtained directly from the diagnostic sample, as VP2 sequencing has been demonstrated to be more sensitive than VP1 sequencing [5]. VP1 sequencing is performed in cases where VP2 typing is unsuccessful, or for specific typing analyses. Non-typable virus isolates are cultivated in two cell-lines

according to WHO guidelines [6] and then characterised by VP1 and VP2 sequencing. For this study, all samples that were positive for EV71 in diagnostic PCR were tested by VP1/VP2 sequencing. VP1 typing and sequencing was applied to comply with international EV characterisation standards. The sample materials for EV71 C4 positive patients are further described (results section).

RNA was extracted from 200 µl of CSF using the QiaCube with the Qia AMP DNA Blood Mini Kit (QIAGEN Nordic, Copenhagen, Denmark), or from 200 µl of other sample material (such as faeces, swabs, biopsies) using the MagNa pure LC robot with the total nucleic acids kit (Roche Diagnostics A/S, Hvidovre, Denmark). Diagnostic PCR for EV was conducted as described previously [7]. 5 µl of the extraction was used as template for PCR amplifying part of the VP1 and VP2 gene, respectively in semi-nested PCR [5]. cDNA synthesis and first round PCR was carried out using a OneStep reverse transcription (RT)-PCR kit (QIAGEN Nordic, Copenhagen, Denmark), and second round amplification was carried out, producing a VP1 amplicon of 350 to 400 basepairs and a VP2 PCR amplicon of 368 basepairs.

Prior to sequencing, PCR products were purified using exo-SAP IT (GE Healthcare, Buckinghamshire, UK). Purified PCR products were sequenced in both directions. Phylogenetic analysis based on the VP1

FIGURE 1

Distribution of enterovirus genotypes, Denmark, 2009–2013 (n=938)^a

Genotypes for which 10 or more viruses were detected are shown. Genotypes for which fewer than 10 viruses were detected are grouped together according to which enterovirus (EV) species they belong to, i.e. EVA, EVB, EVC, and EVD. The viruses which are grouped according to species are shown immediately after the last genotype of that species in the chart.

^a A total of 938 strains were characterised from 913 individuals. Of the 938 strains, 25 represent either co-infections with two different genotypes, or a subsequent infection with a different genotype.

FIGURE 2

Distribution of enterovirus 71 subgenotypes C1, C2, and C4, Denmark, 2009–2013 (n=63)

sequences was carried out by maximum likelihood and the Kimura 2-parameter model with discrete gamma distribution and invariable sites, and 1,000 bootstrap replications using the Molecular Evolutionary Genetics Analysis (MEGA)5 software package [8]. VP1 sequence data from 23 of the 34 EV71 subgenotype C4 samples were of sufficient quality and length to be included in the phylogenetic analysis (sequence length 178–305 nucleotides). All sequences from this study have been submitted to GenBank. Two subgenotype C1 strains and 11 subgenotype C2 strains were also included, as were all publically available (GenBank) European EV71 C4 sequences. The analysis was supplemented with reference sequences obtained from GenBank representing EV71 subgenotypes A, B1 to B5, and C1 to C5 (including C4a and C4b).

Results

In the period from 1 January 2009 to 31 December, 2013 a total of 7,879 samples from 5,611 individuals were received at the Department of Microbiology Diagnostic and Virology at SSI for EV diagnostics. Of these, 984 (12%) samples from 779 (14%) different individuals were positive for EV. In the same period 544 EV positive samples from 427 individuals were received for our national surveillance, thus totalling 1,447 positive samples from 1,143 individuals. 938 EV samples from 913 individuals were successfully genotyped. In 25 individuals more than one EV genotype was detected. A total of 41 different genotypes were identified, the most prevalent being Eo6 (n=141/913, 15%), E30 (n=114/913, 12%), CAo6 (n=96/913, 11%), and EV71 (n=63/913, 7%) (Figure 1).

EV71 was detected throughout the entire period, however there was a marked shift in the subgenotype, from C1 (n=3) and C2 (n=23), being found mainly between 2009 and 2011 with additionally C2 in 2012 (n=1) and 2013 (n=2), to subgenotype C4 being found primarily in 2012 (n=30) but also in 2013 (n=4) (Figure 2). The 34 EV71 C4 infected individuals were unevenly distributed

with regard to sex, as 22 of the 34 cases were males. With regards to age, the majority of infected individuals were young children, with 30 of the 34 C4 cases ≤5 years-old, and 16 <1 year-old.

With regard to the severity of symptoms, patients infected with the C4 subgenotype showed comparable symptoms to patients infected with subgenotypes C1 and/or C2 (Table 1 and 2). Nineteen of the 34 C4 patients had cerebral or sepsis-like symptoms. Additional symptoms among the EV71 C4 infected cases were gastroenteritis (n=7), and HFMD (n=4). EV71 C4 was detected primarily from stool samples (n=19/34, Table 1). Except for the single clustering of EV71 C4 cases in Funen during the months of July to December of 2012 (n=12), most single cases appear sporadically throughout the study period and from all five major geographical regions of Denmark (Table 1). Of the 12 clustered cases from Funen, 10 were admitted to the central hospital and one case was referred to this hospital from a nearby provincial hospital. The age range was 0 to 40 years (median: 2 years), with an uneven sex distribution of nine males, and three females.

The phylogenetic analysis revealed one major C4 lineage, containing all of the C4 strains reported in this study (Figure 3). These were determined to belong to the C4a lineage from Asia. Previously reported C4 strains from Europe belong to the C4b lineage [2–4].

Discussion

This study reports the finding of a new EV71 C4a subgenotype, detected in Denmark for the first time in the spring of 2012. As of December 2013 a further 33 EV71 C4 cases have been detected, the majority in infants with moderate to severe symptoms. EV71 C4 cases from Austria and Hungary were also found to be associated with severe symptoms such as meningitis and acute flaccid paralysis [2,3]. In Denmark, study material is based on cases referred for either diagnostic purposes, or submitted to the National WHO Polio Reference Laboratory at SSI, as part of the national EV surveillance. As a consequence, the detection of mild and/or asymptomatic cases of EV71 infection in the Danish population is not complete, and we can therefore not conclude that EV71 C4 is always associated with severe symptomatology. Only 6/63 EV71 cases were associated with HFMD. There is no specific surveillance for HFMD in Denmark, so the actual level of mild cases of EV71 in circulation may be underestimated.

The EV71 C4 strains identified in Denmark shared a surprisingly high sequence similarity with an EV71 C4a epidemic strain from China, 2008 (EU913466, Figure 3) [9–11]. So far, the relatively severe presentation, although with no fatalities, of 19 of the 34 EV71 C4 cases, with a temporal-spatial clustering of nine of the meningitis/encephalitis cases in Funen during the second half of 2012, suggests that the Danish emerging C4 strain has the same potential for a high transmission

TABLE 1

Characteristics of enterovirus 71 C4 infected patients, Denmark, 2009–2013 (n=34 patients)

Patient number	Age in Years	Sex	Date of sampling	Geographical region	Primary symptoms	Sample material
1	0	M	11-02-2012	Funen	Meningitis	Faecal swab
2	0	M	22-05-2012	Jutland	Meningitis	Stool
3	2	M	06-07-2012	Funen	Gastroenteritis	Swab
4	4	M	09-07-2012	Funen	Meningitis	Faecal swab
5	3	F	23-07-2012	Jutland	Encephalitis	Stool
6	0	F	01-08-2012	Jutland	Gastroenteritis	Stool
7	0	F	22-08-2012	Funen	Sepsis-like syndrome	Stool
8	40	M	30-08-2012	Funen	Meningitis	CSF
9	2	M	04-09-2012	Funen	Meningitis	Stool
10	0	F	11-09-2012	Zealand	Fever	Stool
11	1	M	13-09-2012	Zealand	Unknown	Fluid
12	6	M	27-09-2012	Jutland	HFMD	Swab
13	29	M	11-10-2012	Jutland	Respiratory symptoms	Swab
14	0	F	14-10-2012	Jutland	Gastroenteritis	Stool
15	0	M	17-10-2012	Funen	Encephalitis	Swab
16	3	M	26-10-2012	Funen	Meningitis	Swab
17	0	M	26-10-2012	Jutland	Meningitis	Stool
18	1	M	30-10-2012	Funen	Encephalitis	Swab
19	0	M	04-11-2012	Funen	Gastroenteritis	Stool
20	0	M	05-11-2012	Funen	Meningitis	Stool
21	0	M	07-11-2012	Zealand	Sepsis-like syndrome	Stool
22	0	F	08-11-2012	Zealand	Sepsis-like syndrome	Stool
23	0	M	09-11-2012	Funen	Meningitis	Stool
24	4	M	16-11-2012	Jutland	Meningitis	Stool
25	1	F	25-11-2012	Jutland	Fever	Stool
26	1	F	28-11-2012	Jutland	Gastroenteritis	Stool
27	33	F	06-12-2012	Zealand	HFMD	Vesicle fluid
28	2	M	07-12-2012	Zealand	HFMD	Swab
29	3	M	11-12-2012	Jutland	Gastroenteritis	Stool
30	4	F	18-12-2012	Funen	Meningitis	Swab
31	0	M	08-01-2013	Jutland	Gastroenteritis	Stool
32	2	F	16-07-2013	Jutland	Meningitis+HFMD	Faecal swab
33	0	M	22-08-2013	Zealand	Meningitis	CSF
34	0	F	03-10-2013	Jutland	Fever	Stool

CSF: cerebrospinal fluid; HFMD: hand foot and mouth disease; M: male; F: female.

rate and high pathogenicity as described previously for Asian EV71 C strains, including EV71 C4 [9–13].

Other EV71 C subgenotypes, namely C1 and C2, have previously been identified as occurring sporadically in Denmark throughout a four year study period (2005 to 2008), implying simultaneous circulation of these lineages without genetic selection of either strain based on VP2 sequences [14]. However, only one EV71 C2 case was identified during 2012 and two in 2013, suggesting that the introduction of C4 within the population of Denmark might have a suppressive impact on the circulation of other EV71 C subgenotypes. Furthermore,

the number of EV71 positive samples in 2012 (n=31) is in itself notable, as a total of only 29 EV71 cases were identified throughout the previous four-year study period [11]. It will be interesting to follow the emergence of C4, and see whether it will follow the typical trend of the other EV71 subgenotypes with only limited evolutionary change within its two lineages (C4a and C4b) over time, or whether this subgenotype will continue to dominate the future EV seasons and give rise to outbreaks of severe disease in Europe, as the C4s are known for in parts of Asia. The increasing number of EV71 C4 identified during the 2009 to 2013 surveillance period, and the initial clustering of 11 cases within one

TABLE 2

Main clinical presentation of enterovirus (EV)71 C4 and EV71 C1 and EV71 C2 patients, Denmark, 2009–2013 (n=63)

Symptom	C4 patients	C2+C1 patients
Meningitis	13	12
Gastroenteritis	7	7
HFMD	4	2
Encephalitis	3	2
Sepsis-like syndrome	3	1
Respiratory symptoms	1	1
Fever	3	6
Myoclonus	0	1
Unknown	1	1
Total number of patients	34 ^a	29 ^a

HFMD: hand foot and mouth disease.

^a Some patients presented more than one symptom so the total numbers of patients are not equal to the sum of the numbers in the respective columns.

hospital during the second half of 2012, suggest that this subgenotype initially gave rise to a smaller localised outbreak and is potentially now establishing itself in this northern European population. In the recent 'perspective' from the Global Disease Detection Operations Center at the United States Centers for Disease Control and Prevention (CDC), EV71 was considered to be one of the top-five global infectious disease threats to watch, due to its propensity to cause large outbreaks and severe, life threatening, neurologic disease [15].

In conclusion, the circulation of EV71 subgenotype C4a in Denmark has been established. Based on observations using a wide range of different samples from patients with a broad range of EV symptoms, this subgenotype was found to coincide with severe disease, as were the other EV71 subgenotypes C1 and C2, detected in Denmark during the study period. EV surveillance of high quality and high sample volume is needed to closely monitor the continued emergence of EV71 C4 in the European population over the coming years to establish the pathogenicity and virulence of this subgenotype. A broader emergence of EV71 within Europe might potentially widen the focus of the current development of EV71 vaccines for targeted use in Asia, to a potential future benefit in Europe as well [16,17].

FIGURE 3

Phylogenetic analysis of viral protein 1 sequences from Danish enterovirus 71 strains, Denmark, 2009–2013

Danish enterovirus (EV)71 strains are represented with a black square. The sequence identifiers for the sequences obtained in this study are made up of a two figure prefix to denote the year of sample collection, followed by our internal sample number. Subgenotype C reference sequences were downloaded from GenBank (EV71 C1 accession number: DQ341360; EV71 C2: AF136379; EV71 C3: DQ341355; EV71 C4a: EU913466; EV71 C4b: EU547500; EV71 C5: EU527983). EV71 subgenotypes A (EV71A: ETU22521) and B (B1: AB482183; B2: ETU22522; B3: EU364841; B4: DQ341365; B5: DQ341362) were included to root the tree. Previously identified European C4 strains were also included in the analysis. The scale bar represents the number of nucleotide substitutions per site.

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Conflict of interest

None declared.

Authors' contributions

TKF: conceptualised the study, drafted the manuscript and headed the investigation; AYN: participated in the molecular and phylogenetic analyses, and has participated in the writing of the manuscript; TVS: collected clinical information on the most important samples; PHA: participated in the national enterovirus surveillance and in the registration of clinical symptoms; BA: conducted the laboratory characterisation of EV; SOI: headed the laboratory characterisation of EV and the phylogenetic analyses, and has participated in the writing of the manuscript.

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