Rapid communications

Dengue virus serotype 4 and chikungunya virus coinfection in a traveller returning from Luanda, Angola, January 2014

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Citation style for this article:

A concurrent dengue virus serotype 4 and chikungunya virus infection was detected in a woman in her early 50s returning to Portugal from Luanda, Angola, in January 2014. The clinical, laboratory and molecular findings, involving phylogenetic analyses of partial viral genomic sequences amplified by RT-PCR, are described. Although the circulation of both dengue and chikungunya viruses in Angola has been previously reported, to our knowledge this is the first time coinfection with both viruses has been detected there.

Detection of coinfection

Here we report the simultaneous detection of chikungunya virus (CHIKV) and dengue virus (DENV) genomes in the peripheral blood of a traveller who returned from Luanda, Angola, to Portugal in January 2014.

The traveller, a woman in her early 50s, was born and raised in Angola and has lived in Lisbon, Portugal, since the early 1990s. She stayed in Luanda from mid-December 2013 to early January 2014 at her family’s place of residence. There were a large number of mosquitoes in the garden and the patient was repeatedly bitten during her stay.

The patient reported feeling unwell in early January, two days before her return to Portugal. Her condition worsened during the flight, and in the next few days she had high fever (up to 39.5 °C), severe arthralgia, myalgia, prostration and abdominal pain. Three days after her return, she went to the emergency department of a hospital: a malaria blood smear was negative and among a range of laboratory tests (including coagulation speed and levels of glucose, creatinine, bilirubin, aspartate transaminase (AST), alanine transaminase (ALT), lactate dehydrogenase (LDH), sodium, potassium, chloride ions and C-reactive protein), the only abnormal findings were a mildly low platelet count (139 × 10^9/L; norm: 150–400 × 10^9/L) and mild leucopenia (2.9 × 10^9/L; norm: 4–10 × 10^9/L). The following day, she went to a hospital specialised in tropical diseases, where photophobia was detected. Further tests were carried out (described below). An arbovirus infection was suspected as the malaria blood smear was persistently negative.

Four days later, the fever had subsided and her condition improved progressively over the next two to three weeks. The patient did not have a rash, conjunctivitis or other clinical signs of a complicated dengue infection (DENV infection with haemorrhage); indeed, she had no other abnormal clinical signs at all during the course of her illness. To the best of her knowledge, none of her family or neighbours in Luanda experienced a similar illness.

Laboratory findings

Four days after her return from Luanda, DENV non-structural (NS) protein 1 and anti-CHIKV IgM were detected (through the use of SD BIOLINE Dengue Duo NS1 Ag + Ab Combo and SD Bioline Chikungunya IgM), while DENV-specific IgM and IgG were not detected. Two days later, the same tests were performed: anti-CHIKV IgM and DENV-specific IgM and IgG were detected, but DENV NS1 was not. Using RNA extracted from the blood sample where NS1 had been found, detection of the viral genomes was carried out either by a nested RT-PCR as previously described [1,2] or by using primers that target the virus packaging sequence [3]. The sizes of the amplicons obtained were compatible with the presence of both DENV4 (approximately 390 bp, covering the C-prM region) and CHIKV (approximately 350 bp, in the NS2 coding region).
Additional molecular confirmation was obtained by performing phylogenetic analyses of the sequence of both amplicons (deposited in the GenBank/European Molecular Biology Laboratory (EMBL)/DNA DataBank of Japan (DDBJ) databases under accession numbers AB908053 and AB908054) using the using GTR+Γ+I model [4]. The DENV sequence obtained clearly clustered with DENV4 reference strains (Figure 1), while the CHIKV sequence segregated with those included in the Central/Eastern/Southern African genotype (Figure 2). Despite the presence of both viral genomes in the same blood sample, the viraemia dropped rapidly below the detection level, as both DENV and CHIKV RNA could not be detected in blood collected 48 hours later.

Background

Dengue has developed into a worldwide public health problem, especially over the last 50 years [5,6]. More recently, the impact of other arboviruses on human health has followed a similar trend [7]. This is true for CHIKV, which, since 2004, has been an emerging pathogen, causing large outbreaks in many islands in the Indian Ocean and in the Indian subcontinent, where, in 2005-2006 alone, well over a million cases of CHIKV infection were reported from different states [8].

The majority of DENV infections occur in the Asia–Pacific and Americas–Caribbean regions [5], while CHIKV is endemic to countries in Africa and Asia [9]. In Africa, the epidemiology and public health impact of both viruses is far from clear, but the wide geographical distribution of their primary vectors (Aedes aegypti and Aedes albopictus), rapid human population growth, unplanned urbanisation, and increased international travel make their transmission likely [10,11]. Moreover, as the clinical features of DENV and CHIKV are similar, CHIKV infections usually go undiagnosed in areas where DENV circulates [11]. Furthermore, where malaria is also endemic and the majority of febrile illnesses are diagnosed as such, often without laboratory confirmation, both viral infections may go undetected [12].

Although CHIKV/DENV coinfections were first reported in India in 1967 [13] and later confirmed in Sri Lanka (2008), Malaysia (2010) and Gabon (2007) [14-16], these coinfections are rarely notified.

Discussion

Serological reports from the 1960s [17], the detection of DENV in travellers returning from Angola in the 1980s [10], and the detection of DENV1 and DENV2 in travellers in the 1980s and in 1999–2002 [10,18] suggest endemic DENV activity in Angola. As far as CHIKV is concerned, the situation is a lot less clear. However, serological studies from the 1960s not only identified the presence of anti-CHIKV neutralising antibodies in the north of the country, but also allowed the isolation of two strains from a viraemic individual and wild-caught mosquitoes during an outbreak of Kâtolu Tôlu (Kimbundu dialect for ‘break-bone disease’), a dengue-like disease caused by the CHIKV, which occurred in Luanda in 1970 [19].

The detection of DENV4 in the recent traveller is of interest, given that on 1 April 2013, the Angolan health authorities reported a dengue outbreak in the country [20], which was later shown to have been caused by DENV1 [21], and the current description of DENV4 in

![Figure 1](image-url)

Maximum likelihood phylogenetic tree analysis of dengue virus (DENV) serotypes 1–4 C-prM sequences.

The tree was constructed using the using the GTR+Γ+I model [4]. The amplicon isolated from the patient is shown in bold. Reference strains, downloaded from public databases, are identified by strain name and accession number (DENV4) or simply by viral serotype and accession number (DENV1–3). The numbers at specific branches indicate bootstrap values (only values ≥77% are indicated).
Luanda may indicate the circulation of multiple DENV subtypes in the country.

Although clinical examination of CHIKV/DENV coinfected patients has not yet allowed the identification of specific or severe symptoms, such observations should be interpreted with caution in view of the limited number of clinical and biological investigations reported. Our findings may add to the recognition of CHIKV/DENV coinfections and suggest that tests to detect the presence of both viruses should be carried out in individuals showing clinical signs of an infection with either CHIKV or DENV.

Conflict of interest
None declared.

Authors’ contributions
Ricardo Parreira: molecular analyses and manuscript writing. Ângela Mendes: molecular analyses. Jaime Nina: clinical diagnosis and manuscript writing. Antónia Constantino: clinical diagnosis and manuscript writing. Sónia Centeno-Lima and Daniela Portugal Calisto: laboratory diagnosis and manuscript writing.
References


